A brief communication of patients with homozygous C282Y mutation-related hereditary hemochromatosis

D Hale Gokcan¹, D Didem Kuru Oz², Emin Bodakci¹, Esra Tunc³, Ramazan Idilman^{1,3}

¹Department of Gastroenterology, Ankara University School of Medicine, Ankara, Turkiye; ²Department of Radiology, Ankara University School of Medicine, Ankara, Turkiye; ³Institute of Hepatology, Ankara University School of Medicine, Ankara, Turkiye

Abstract

Hereditary hemochromatosis (HH) is an autosomal recessive inherited iron-loading disorder and is characterized by chronic hepatitis, cirrhosis, diabetes, and bronze skin. The hemochromatosis gene (C282Y homozygosity)-related hemochromatosis is the most common form of HH. The prevalence of HH is varied. Here, we defined six cases with C282Y homozygosity-related HH in a single center in Turkiye.

Keywords: C282 Y homozygous mutation; hereditary hemochromatosis; HFE gene.

Introduction

Hereditary hemochromatosis (HH) is an autosomal recessive disorder of iron metabolism that is characterized by inadequate hepcidin synthesis, increased intestinal iron absorption, and iron release from macrophages, leading to progressive parenchymal iron accumulation with the potential for multiorgan damage. Excess iron is deposited in the liver, pancreas, heart, endocrine glands, and joints.^[1] HH mainly manifests in the liver and can result in fibrosis, cirrhosis, and hepatocellular carcinoma (HCC).^[2] Homozygotes for the hemochromatosis gene (HFE) C282Y is the most common genotype in HH and is seen in 80% to 95% of cases of typical HH among European individuals.^[3,4] In Turkiye, HFE-associated HH is rarely seen.^[5] Moreover, this brief study represents the largest series reporting HFE-associated hemochromatosis due to C282Y homozygosity in a single center of Turkiye.

Materials and Methods

We investigated hereditary HFE-associated hemochromatosis due to C282Y homozygosity in patients followed by the Liver Disease Outpatient Clinic, Ankara University School of Medicine, between Janu-

How to cite this article: Gokcan H, Kuru Oz D, Bodakci E, Tunc E, Idilman R. A brief communication of patients with homozygous C282Y mutation-related hereditary hemochromatosis. Hepatology Forum 2024; 5(3):161–164.

Received: May 30, 2024; Accepted: June 13, 2024; Available online: July 02, 2024

Corresponding author: Hale Gokcan; Ankara Universitesi Tip Fakultesi, Gastroenteroloji Anabilim Dali, Ankara, Turkiye

Phone: +90 312 595 70 40; e-mail: halesumer@yahoo.com

OPEN ACCESS This work is licensed under a Creative Commons Attribution-NonCommercial 4.0 International License

Hepatology Forum - Available online at www.hepatologyforum.org



ary 2015 and December 2023. The diagnosis of hemochromatosis was based on clinical features, biochemical tests, including transferrin saturation (TS) (normal range: 13-45%) and serum ferritin level (normal range: female 13-150 ng/mL, male 30-400 ng/mL), imaging methods (magnetic resonance imaging [MRI]), genetic screening, family history, and histological evaluation when available. Secondary causes of iron overload, such as hematological disorders, history of multiple transfusions, parenteral iron therapy, alcohol-related liver disease (ALD), and metabolic dysfunction-associated steatotic liver disease (MASLD), were also reviewed for differential diagnosis.

HEPATOLOGY FORUM

doi: 10.14744/hf.2024.2024.0020

Genomic DNA was isolated from a whole blood sample in an EDTA tube using a genomic DNA isolation kit (Roche, Mannheim, Germany). The DNA region containing the HFE gene was amplified by PCR using appropriate primers. The amplified PCR products underwent sequencing using a genetic analyzer (BigDye terminator kit, ABI 310 Genetic Analyzer, Mannheim, Germany).

MRI was performed on 1.5 Tesla MRI devices (Optima 450w, GE Healthcare, and Aera, Siemens Healthcare). The liver-to-muscle signal intensity ratio method, a calculation tool from the University of Rennes in France, was used to estimate liver iron concentration in a subset of patients.^[6] The normal liver iron concentration (LIC) value is less than 36 µmol Fe/g. In HH, it is greater than 80 µmol Fe/g.^[7] For the remaining patients, quantitative T2* and its reciprocal R2* (R2*=1000/T2*) maps were obtained using a multi-echo spoiled gradient-echo single-breath-hold MR sequence. In healthy individuals, the T2* value is above 14 ms, and the R2* value is below 70 s^{-1.[8]}

MR-based fat quantification [proton density fat fraction (PDFF)] was mainly measured using a chemical shift (CS)-based MRI method known as multi-echo Dixon for Siemens. This method employs six echoes and can generate a comprehensive PDFF map of the entire liver. An FF threshold of 6.3% was accepted as the upper normal limit.^[9]

Results

Six patients had been diagnosed with C282Y homozygotes HH: four were male and two female. The mean age at diagnosis was 48.6 ± 16.5 years. Their median TS and serum ferritin levels were 77.5% (range 43-82) and 1757.5 ng/mL (range 256-5264), respectively. The characteristics of the patients with HH are given in Table 1.

Cases 1 and 6 were siblings. Cases 1, 2, 3, and 6, male patients, represented a clinical expression of HH. However, the female cases (Cases 4 and 5) had no phenotypic manifestation of hemochromatosis. Two patients (Cases 2 and 3) represented cirrhosis: one had hepatocellular carcinoma (HCC), three patients (Cases 1, 4, 6) with hepatosteatosis (2

Tal	ole 1. Chara	cteristi	cs of pa	atients with C282	Y homozygotes h	emochromatosis			
С	Age at diagnosis	Sex	DD	Hepatic manifestation at diagnosis	Extrahepatic manifestation at diagnosis	TS (normal range: 13-45%) / Ferritin (normal range: female 13-150 ng/ mL, male 30-400 ng/mL)	MRI at diagnosis	Liver biopsy at diagnosis	Family history
1	25	Μ	2021	Hepatosteatosis		76/1442	Severe hepatosteatosis (Dixon FF= 24%) Mild-moderate iron accumulation ($R2^*=245 s^{-1}$ $T2^*=4.5 ms$)	60% macrovesicular steatosis, HIC: 1600 µg/gram dry weight HII: 1.6	Sibling of case 6
2	52	Μ	2015	Cirrhosis	Diabetes mellitus Hypopituitarism Libido loss Hypothyroidism Heart failure Atrial fibrillation	79/3332	Severe iron accumulation (LIC at PD sequence=350 µmol/g)	Grade 4 iron staining	
3	67	М	2016	Cirrhosis with HCC	Bronze diabetes arthropathy	72/5264	N/A	N/A	
4	52	F	2018	Hepatosteatosis		82/256	N/A	N/A	Sister
5	63	F	2014	Chronic HBV infection		43/1501	No iron accumulation LIC at GRE sequence=30 µmol/g	N/A	
6	33	М	2021	ALD		81/2014	Severe hepatosteatosis (DixonFF=20% Moderate iron accumulation (R2*=241 s ⁻¹ T2*=3.8 ms)	40% macrovesicular steatosis, HIC: 2500 μg/gram HII: 1.35	Brother of case 6

C: Case; DD: Diagnosis date; M: Male; F: Female; TS: Transferrin saturation; MRI: Magnetic resonance imaging; FF: Fat fraction; HIC: Hepatic iron concentration; HII: Hepatic Iron Indeks; LIC: Liver iron concentration; HCC: Hepatocelular carcinoma; HBV: Hepatitis B virus; ALD: Alcoholic liver disease.

MASLD, 1 ALD), and the remaining one patient (Case 5) with chronic hepatitis B virus (HBV) infection.

Discussion

Cases 2 and 3 represented typical clinical expressions of HH. Case 2 had diabetes mellitus, hypopituitarism (hypothyroidism, libido loss), congestive heart failure (due to cardiomyopathy), and atrial fibrillation. Case 3 had bronze diabetes and arthropathy.

Case 4 had a family history of HH, and her sister was diagnosed with HH in another center. Her TS and ferritin levels were 82% and 256 ng/ mL, respectively. Case 5, a female patient, had a TS of 43% with high ferritin levels (1501 ng/mL).

On the other hand, the C282Y heterozygous (C282Y/N) mutation was found in five patients (Cases 7, 8, 9, 10, and 11), and the compound C282Y heterozygous (C282Y/H63D) mutation was found in one patient (Case 12). Four patients (Cases 7, 8, 10, 12) were represented with hepatosteatosis (2 MASLD, 2 ALD). Case 7 had chronic hepatitis C, and he underwent renal transplantation. Case 9 was diagnosed with polycythemia vera, and Case 11 with ALD. Case 10 and Case 11 were the parents of Case 1 and Case 6. Demographic and laboratory data of patients based on HFE gene mutations are presented in Table 2.

This study determined the largest series of HFE-associated hemochromatosis due to C282Y homozygosity in a single center in Turkiye. The frequency of C282Y homozygosity differs across Europe depending on the geographical region. It is most common in individuals of Northern European descent, particularly in Ireland and Scandinavia. [3,4,10] Patients with C282Y homozygosity are at risk of developing hemochromatosis, but the disease penetrance depends on age and gender. The prevalence of the disease increases with age, and male gender is predominant.^[1,11] HFE-related HH is a multifactorial disease characterized by stepwise disease progression from biochemical test abnormality to organ damage. The altered HFE gene plays an essential role in disease progress. However, host-related factors such as age, gender, other genes, and acquired factors (diet and alcohol consumption) also play important roles in the phenotypic penetrance of the genetic defect.^[12] These factors may lead to biochemical abnormalities, symptoms, and signs or organ damage with/without overt organ failure. Many patients are asymptomatic and have been followed up for many years, while

Table 2. Demogra	phic and	laboratory	y data of	patients	based on	the HFE	gene mutation							
				C282Y h	omozygoı	Sľ			C282Y he	terozygo	us/ C282'	Y compor	und hete	rozygous
Case	-	7	m	4	Q	9	Mean±SD /Median (Range)	7	œ	G	6	1	12	Mean±SD ∕Median (Range)
Age	25	52	67	52	63	33	48.6±16.5	42	55	28	60	61	73	53.1±15.8
Gender	Σ	Σ	Σ	ш	ш	Σ		ш	Σ	Σ	ш	Σ	Σ	
Hemoglobin (g/L)	15.7	13.0	15.2	14.0	14.0	14,7	14.4±0.96	12.3	14.5	17.4	14.3	14.1	16.1	14.7±1.76
AST (U/L)	40	52	104	17	19	67	46 (17–104)	33	68	16	27	23	33	30 (16–68)
ALT (U/L)	106	31	94	20	16	143	62.5 (16-143)	53	60	1	34	25	35	34.5 (11–60)
GGT (U/L)	37	175	85	22	13	34	35.5 (13–175)	16	226	10	21	23	81	22 (10–226)
Albumin(g/dL)	48.4	36.0	33.3	43.0	42.0	47.0	41.5±6.0	46.7	47.6	51.6	47.9	46.3	47.0	47.8±1.9
INR	1.12	1.23	0.98	0.95	0.99	0.98	1.04±0.1	0.84	0.84	1.03	1.02	1.1	0.97	0.96±0.1
Bilirubin(mg/dL)	0.34	1.30	0.88	1.10	0.56	0.6	0.79±0.36	1.23	0.98	1.91	0.5	0.5	0.81	0.98±0.53
TS (%)	76	79	72	82	43	81	77.5 (43–82)	71	61	52	43	38	40	47.5 (38–71)
Ferritin (ng/mL)	1442	3332	5264	256	1501	2014	1757.5 (256–5264)	952	1106	36	262	76	580	421 (36–1106)
SD: Standard deviation;	M: Male; F:	Female; AST	⁻: Aspartat ar	ninotransfen	ase; ALT: Al∉	anine aminoti	ransferase; GGT: Gamma glu	ıtamil transfe	erase; INR: Ir	nternational	normalized	ratio; TS: Tr	ansferrin :	aturation.

some patients present with cirrhosis or bronze diabetes.^[13] In this series, four of six patients with C282Y homozygotes were male, and they had a clinical expression of hemochromatosis. However, two female patients with C282Y homozygosity were asymptomatic and had no significant iron overload.

The clinical impact of C282Y heterozygotes and compound C282Y/ H63D heterozygotes appears limited. The patients having these mutations have slightly increased serum iron parameters and hepatic iron load and seem to be predisposed to the expression of the disease.^[14,15] In these cases, secondary causes of iron overload, such as MASLD, alcohol consumption, iron-loading anemias, parenteral iron administration, ferroportin disease, aceruloplasminemia, or atransferrinemia should also be investigated.^[4,11] In the present report, we found five patients with C282Y heterozygotes and one with compound C282Y/H63D heterozygotes. Hepatosteatosis is the most common clinical expression in these cases.

The initial step in the diagnosis of hemochromatosis is to suspect the disorder in patients with unexplained liver injury test abnormalities, bronze diabetes, arthralgia, cardiomyopathy, or hypogonadism. Family history, especially first-degree relatives, should be investigated. High transferrin saturation and serum ferritin levels can be used to screen and diagnose HFE-related HH. These tests lead to occasionally missed diagnoses and some false-positive results. HFE mutation analysis is typically the initial genetic test conducted in individuals with hemochromatosis, especially in European countries due to a high prevalence of HH.^[1,11] In Turkive, the HFE gene mutation analysis should not be considered an initial test in the diagnosis of hemochromatosis because of its low prevalence. MRI techniques are available for quantifying liver iron concentration and have shown a good correlation with liver biopsy.^[16] Confounder-corrected R2*-based liver iron content (LIC) quantification is the most clinically useful method, with the strongest supporting evidence for providing a precise and reliable measurement of LIC. This technique is commonly available in many MRI scanners. Although liver biopsy is currently the preferred diagnostic and prognostic indicator method for iron overload disorders, in countries with a low prevalence of HH, it is feasible to evaluate organ iron accumulation with MRI at first to exclude the disease.

Conclusion

In conclusion, hemochromatosis has been a life-threatening disease with high morbidity and mortality in some cases. Patient and physician awareness of hemochromatosis is the first step toward the successful management of the disease.

Author Contributions: Concept – HG; Design – HG; Supervision – HG, RI; Fundings – HG, EB; Materials – HG, EB, ET; Data Collection and/or Processing – HG, DKO, EB, ET; Analysis and/or Interpretation – HG, RI, DKO, ET; Literature Search – HG, RI; Writing – HG, RI; Critical Reviews – HG, RI.

Conflict of Interest: The authors have no conflict of interest to declare.

Informed Consent: Written informed consent was ontained from the patients for the publication of the brief report and the accompanying images.

Use of AI for Writing Assistance: Not declared.

Financial Disclosure: The authors declared that this study has received no financial support.

Peer-review: Externally peer-reviewed.

References

- Liu Yin J, Cussen C, Harrington C, Foskett P, Raja K, Ala A. Guideline Review: European Association for the Study of Liver (EASL) Clinical Practice Guidelines on Haemochromatosis. J Clin Exp Hepatol 2023;13(4):649-655.
- Atkins JL, Pilling LC, Masoli JAH, Kuo CL, Shearman JD, Adams PC, et al. Association of hemochromatosis HFE p.C282Y homozygosity with hepatic malignancy. JAMA 2020;324(20):2048-2057. [CrossRef]
- Porto G, Brissot P, Swinkels DW, Zoller H, Kamarainen O, Patton S, et al. EMQN best practice guidelines for the molecular genetic diagnosis of hereditary hemochromatosis (HH). Eur J Hum Genet 2016;24(4):479-495.
- Crawford DHG, Ramm GA, Bridle KR, Nicoll AJ, Delatycki MB, Olynyk JK. Clinical practice guidelines on hemochromatosis: Asian Pacific Association for the Study of the Liver. Hepatol Int 2023;17(3):522-541. [CrossRef]
- Simsek H, Balaban YH, Yilmaz E, Sumer H, Buyukasik Y, Cengiz C, et al. Mutations of the HFE gene among Turkish hereditary haemochromatosis patients. Ann Hematol 2005;84(10):646-649. [CrossRef]
- Ruefer A, Bapst C, Benz R, Bremerich J, Cantoni N, Infanti L, et al. Role of liver magnetic resonance imaging in hyperferritinaemia and the diagnosis of iron overload. Swiss Med Wkly 2017;147: w14550. [CrossRef]
- European Association For The Study Of The Liver. EASL clinical practice guidelines for HFE hemochromatosis. J Hepatol 2010;53(1):3-22. [CrossRef]
- Lidén M, Adrian D, Widell J, Uggla B, Thunberg P. Quantitative T2* imaging of iron overload in a non-dedicated center - Normal variation, repeatability and reader variation. Eur J Radiol Open. 2021 May 24;8:100357. [CrossRef]

- Tang A, Tan J, Sun M, Hamilton G, Bydder M, Wolfson T et al. Nonalcoholic fatty liver disease: MR imaging of liver proton density fat fraction to assess hepatic steatosis. Radiology 2013;267(2):422–431 [CrossRef]
- Merryweather-Clarke AT, Pointon JJ, Jouanolle AM, Rochette J, Robson KJ. Geography of HFE C282Y and H63D mutations. Genet Test 2000;4(2):183-198. [CrossRef]
- European Association for the Study of the Liver. Electronic address: easloffice@easloffice.eu; European Association for the Study of the Liver. EASL Clinical Practice Guidelines on haemochromatosis. J Hepatol 2022;77(2):479-502.
- Anderson GJ, Bardou-Jacquet E. Revisiting hemochromatosis: genetic vs. phenotypic manifestations. Ann Transl Med 2021;9(8):731. [CrossRef]
- Bacon BR, Adams PC, Kowdley KV, Powell LW, Tavill AS. Diagnosis and management of hemochromatosis: 2011 practice guideline by the American Association for the Study of Liver Diseases. Hepatology 2011;54(1):328-343. [CrossRef]
- Neghina AM, Anghel A. Hemochromatosis genotypes and risk of iron overload-a meta-analysis. Ann Epidemiol 2011;21(1):1-14. [CrossRef]
- Pilling LC, Tamosauskaite J, Jones G, Wood AR, Jones L, Kuo CL, et al. Common conditions associated with hereditary haemochromatosis variants: cohort study in UK Biobank. BMJ 2019;364:k5222 [CrossRef]
- Reeder SB, Yokoo T, França M, Hernando D, Alberich-Bayarri Á, Alústiza JM, et al. Quantification of liver iron overload with MRI: Review and guidelines from the ESGAR and SAR. Radiology 2023;307(1):e221856. [CrossRef]