## **Review article**

# Cross talk between genetics and biochemistry in the pathogenesis of hepatocellular carcinoma

Mete Ucdal<sup>1</sup>, 0000-0002-2321-2642

Ayse Burus<sup>2</sup>, 0000-0002-9464-7053

Basak Celtikci<sup>2\*</sup>, 0000-0002-3242-978X

<sup>1</sup>Department of Itnernal Medicine, Faculty of Medicine, Hacettepe University, Ankara, Turkiye

<sup>2</sup>Department of Medical Biochemistry, Hacettepe University, Faculty of Medicine,

Ankara, Turkiye

<sup>#</sup>These authors contributed equally.

# \*Corresponding author: Basak Celtikci

Address: Hacettepe University, Faculty of Medicine, Department of Medical Biochemistry, Sihhiye, Ankara 06100, Türkiye

Tel: +90(312) 305 16 52

Fax: +90(312) 324 58 85

E-mail: <u>basakceltics@gmail.com</u>

Running Title: Hepatocellular Carcinoma Pathogenesis

# Abstract

The liver is a crucial organ in the regulation of metabolism, signaling, and homeostasis. Using recent advanced sequencing technologies, several mutations of genes in major metabolic and signaling pathways have been discovered in the pathogenesis of hepatocellular carcinoma (HCC). These gene signatures alter expression and ultimately affect biochemical pathways via modifying enzyme/protein levels, and result in numerous clinical outcomes related to HCC. It comes with varying forms of genetic and biochemical alterations, associated with carbohydrate, lipid, nucleic acid, and amino acid metabolism, as well as signaling pathways, linked to tumorigenesis. Here, we aim to summarize the main components and mechanisms involved in the progression of HCC with a special focus on the metabolic regulation of key effectors of tumorigenesis, through the crosstalk between genetics and biochemistry. This paper provides an overview of the hepatocellular carcinoma underlying the fundamental effect of gene variations on metabolic and signaling pathways. Since there is still an unmet need for biomarkers and novel therapeutic targets, some of these signature genes or proteins can be used as novel biomarkers for diagnosis, prognosis, and novel potential therapeutic targets for the treatment of HCC.

**Keywords:** Pathogenesis; Hepatocellular Carcinoma; Genetics; Biochemistry; Biomarkers; Therapeutic targets

#### Introduction

Most of the liver diseases, characterized by an extremely dysregulated metabolism in hepatocytes, are characterized by low overall survival, and poor prognosis, and have complex biological processes on gene and protein levels in their pathogenesis. As the liver is the main site of numerous metabolic events in energy metabolism by the regulation of several biomolecules, including carbohydrates, lipids, nucleic acids, and amino acids, the synergy of interconnected pathways should be assessed rather than isolated mechanisms in these diseases. HCC predominantly shows the Warburg effect. In addition, several pathways of lipid metabolism are dysregulated, such as fatty acid synthesis, β-oxidation, and cellular lipidic composition, to support HCC tumorigenesis. Increased aspartate, glutamine, and hydroxyproline correlated with the tumor burden. Altered miRNA expression, DNA copy number variations (CNVs), deletions, and insertions are prevalent genetic occurrences in HCC [1]. Identification of pre-cirrhotic patients who have the potential to progress to HCC is still an unmet dilemma in clinical practice. Novel biomarkers for diagnosis and/or prognosis, and novel potential therapeutic targets for the treatment are urgently needed.

This paper provides an overview of HCC highlighting the critical effect of gene variations on metabolic and signaling pathways, and their cross-talk among each other in the pathogenesis of liver diseases.

# Hepatocarcinogenesis

Our understanding of the molecular pathogenesis of hepatocarcinogenesis has been improved as a result of recent developments in Next Generation Sequencing (NGS) (Whole genome or exome sequencing) and Genome-Wide Association Study (GWAS) technologies. The molecular classification of HCC has so far been based on the genomic, transcriptomic, metabolic, and/or epigenomic profiling of large tumor cohorts employed in multi-omics research [1]. Studies using NGS have shown 40–60 somatic coding alterations per HCC, with 4-6 of them being driver mutations [2]. There are two mutation subtypes: proliferation and nonproliferation [3]. The proliferation class is defined by cell proliferation and survival pathways, such as PI3K-AKT-mTOR, RAS-MAPK, and MET as well as chromosomal instability, TP53 inactivation, FGF19, and CCND1 amplifications [4].

[5]. Nonproliferation classes often have activation of Wnt signaling transducer  $\beta$ -catenin (CTNNB1) and more TERT promoter mutations. These tumors are transcriptionally identical to normal  $\alpha$ -fetoprotein and are associated with alcohol use and HCV infection etiologies [6].

# **Genetic Pathophysiology of HCC**

The most common HCC mutation is affecting the promoter of the enzyme telomerase reverse transcriptase (TERT) [7]. Up to 60% of HCC patients carry TERT promoter mutations [8]. According to the underlying liver disease, the incidence of TERT promoter mutations varied greatly, with the greatest frequency in HCV-related HCC (44%), followed by non-viral (38%) and HBV-related HCC (23%) [9].

Telomerase is a cellular ribonucleoprotein enzyme. It increases cell proliferation through telomere elongation. TERT is a telomerase component responsible for the enzyme's catalytic activity. Its expression is typically strictly restricted in most normal cells, but up regulated in several malignancies [10]. TERT expression is both transcriptionally and post translationally controlled. Some cancer-associated TERT promoter mutations work by producing de novo transcription-factor-binding sites at the transcription level. The two most common TERT promoter mutations are 124C/T and 146C/T, often known as C228T and C250T, respectively. These two mutations produce putative ETS transcription factor-binding sites (TTCC), triggering TERT transcription. mTERT promoter mutations are one of the mutually exclusive telomerase reactivations.[11] Higher TERT expression in advanced HCC might be attributable to causes independent from TERT promoter mutation, or the accumulation of oncogenic pathways with tumor progression [7]. TERT promoter

mutations are commonly linked to CTNNB1 mutations, indicating that the  $\beta$ -catenin pathway and telomerase maintenance are both involved in the development of liver tumors [12]. Novel copper complex (NCB) regulates methionine cycle-induced TERT hypomethylation to promote HCC cell differentiation [13]. The importance of alternate routes leading to telomere synthesis, such as alternative telomere lengthening, must be properly assessed. Other methods of telomerase re-expression also need to be identified [14].

In the combined novel meta-analysis, the new TERT locus rs2242652(A) lowered HCC risk at genome-wide significance (p=6.41×10-9, OR=0.61 (95% CI 0.52 to 0.70). After controlling for sex, age, BMI, and type 2 diabetes, this protective link remained significant (p=7.94×10-5, OR=0.63 (95% CI 0.50 to 0.79). Leucocyte telomere length increased with TERT rs2242652(A) (p=2.12×10-44) [9]. Like this research, they found genome-wide significant correlations between alcohol-related HCC and PNPLA3 and TM6SF2 polymorphisms. They also observed a significant connection with rs708113 in the WNT3A-WNT9A region on chromosome 1q42, which lowered alcohol-related HCC risk. This variation improved immune cell infiltration of tumor tissues and decreased CTNNB1, which often precedes HCC. 39 rs708113 did not prevent HCC patients with persistent HCV or NAFLD [15] [16].

The WNT/ $\beta$ -catenin signaling system is essential for embryogenesis, zonation, and metabolic regulation in the liver. The cytoplasmic  $\beta$ -catenin destruction complex regulates  $\beta$ -catenin stability. Composed of Axin, APC, CK1, and GSK3, the  $\beta$ -catenin destruction complex is responsible for phosphorylating the N-terminal serine and threonine residues of  $\beta$ -catenin. Following recognition by  $\beta$ -TrCP, phosphorylated B-catenin is ubiquitinated

and destroyed through the proteasome pathway. After the Wnt ligand binds to the cell membrane receptors LRP5/6 and Frizzled, the C-terminus of Frizzled undergoes a conformational shift, resulting in the recruitment of Disheveled proteins (DVL). Then, DVL joins Axin through their shared DIX domain, resulting in the dissolution of the  $\beta$ -catenin destruction complex. Thus,  $\beta$ -catenin accumulates in the cytoplasm and is translocated into the nucleus, where it activates the transcription of downstream genes [17]. It is the oncogenic pathway most triggered in HCC by CTNNB1 activating mutations (11-37%) and AXIN1 inactivating mutations (5-15%) or adenomatous polyposis coli gene (APC) (1-2%) [5]. CTNNB1 mutations, which code for  $\beta$ -catenin, are in-frame deletions or substitutions at a hotspot in the domain targeted by the APC/AXIN1/GSK3B inhibitory complex. Tumors with CTNNB1 mutations show a distinct transcriptome profile with upregulation of conventional target genes such as glutamate ammonia ligase (GLUL) and leucine-rich repeat-containing G-protein coupled receptor 5 (LGR5), as well as a distinct histologic pattern with intratumor cholestasis [18].

Numerous nongenetic pathways that activate Wnt/ $\beta$ -catenin have also been found in studies. These include overexpression of the Frizzled membrane receptor (FZD) and Wnt ligands; deregulated expression of microRNAs and long non-coding RNAs that regulate Wnt/ $\beta$ -catenin signaling; and promoter hypermethylation and associated silencing of the secreted Frizzled related protein 1 (SFRP1) gene, a Wnt/ $\beta$ -catenin antagonist [19-21]. In recent years, it has been shown that oncogenic or tumor suppressor noncoding RNAs control the canonical Wnt/ $\beta$ -catenin signaling pathway's major regulatory elements, thereby controlling HCC cell proliferation, invasion, metastasis, and drug sensitivity. MiR-1246 has been identified as a possible factor that promotes HCC tumor growth by

inhibiting the expression of its target ROR. Notably, they found that artificial stimulation of miR-1246 expression or ROR knockdown significantly increases the metastatic ability of HCC in vitro and in vivo via activation of the Wnt/ $\beta$ -catenin pathway and promotion of epithelial-mesenchymal transition (EMT) [22]. Similarly, USP22, a histone-modifying enzyme primarily regulated by miR-329-3p (normally downregulated in HCC), is associated with distant metastasis, poor prognosis, and high recurrence rates in HCC, because it critically modulates tumor cell proliferation, metastasis, DNA repair, and stemness via modulating Wnt/ $\beta$ -catenin pathways [23].

Recently, it was discovered that miR-19a-3p and miR376c-3p may promote the Wnt/βcatenin pathway in HCC cells by targeting SOX6. Furthermore, they discovered that SOX6 may bind to β-catenin, preventing it from dissociating from the transcriptional complex and being translocated to the nucleus, Overall, the results indicated that both miR-19a-3p and miR-376c-3p are significantly expressed in HCC cells and may play a role in HCC development by targeting SOX6 and disrupting the Wnt/β-catenin signaling pathway [24]. At least half of HCC patients with frequent TP53 mutations (12–48%), the tumor suppressor gene more often mutated in cancer, had altered P53 cell cycle pathways. There has not been discovered another recurrent TP53 mutation hotspot other than the R249S mutation linked to AFB1 exposure [25]. The homozygous deletion of CDKN2A (2– 12%), 4562, or RB1 mutations (3–8%) in HCC mostly inactivates the retinoblastoma pathway, which controls progression from the G1 to S phase of the cell cycle. Interestingly, tumors with poor prognosis have higher levels of genetic changes in CDKN2A and RB1, pointing to a role for P21 pathway inactivation in tumor aggressiveness [14]. Receptor tyrosine kinase (RAS-RAF-MAPK) and phosphatidylinositol-3-kinase, Protein kinase B, and mammalian target of rapamycin (PI3K-AKT-mTOR) pathways are generally activated in HCC, due to amplification of areas including FGF19 (5% tumors) and mutations in RPS6KA3 and RSK2 (5-9% instances) [26]. Furthermore, in a subset of HCC, activating mutations of PIK3CA (0-2%) and inactivating mutations of TSC1 or TSC2 (3-8%) result in the activation of the AKT/MTOR signaling. Additionally, homozygous deletion of the tumor suppressor phosphatase and tensin homolog (PTEN), which is a PI3K kinase inhibitor, has been found in 1-3% of HCC [12, 14]. An additional path of AKT/mTOR activation has been proposed: direct upstream activation via the insulin growth factor pathway. Activating mutations in RAS family genes are uncommon in HCC (2%), and inactivating variants in RP6SKA3, which codes for the RAS inhibitor RSK2, were found in (2-9%) of tumors [26]. RSK2 is a recognized negative regulatory loop of RAS signaling that is positioned downstream of MAPK [26]. Inactivation of RSK2 resulted in the release of this negative feedback and constitutive activation of the pathway [12]. Experiment results further imply that prolonged RAS activation may be a cause of sorafenib resistance in HCC [12].

miRNA dysregulation has a well-established role in the PI3K-AKT-mTOR pathway. miR-660-5p, miR-1914, miR-125a-5p and moreover noncoding miRNA activated PI3K/Akt signaling to promote the proliferation and invasion of HCC cells [14]. The major tumor suppressor and oncogenic miRNAs and their related genes and pathways are summarized in Table 1.

 Table 1. Tumor-Suppressor and oncogenic miRNAs and their regulated gene expressions

Tumor Supressor	Regulated Gene Expressions	References
miRNAs		
miR-206	cMET, CCND1 and CDK6	[27]
miR-214	EZH2, β-catenin	[28, 29]
miR-223	STMN1	[30]
miR-302b	AKT2	[31]
miR-339	ZNF689	[32]
miR-15a, miR16 and	WNT3A	[33]
miR-107	•	
miR-375	YAP and AEG-1	[34]
miR-424-5p	TRIM 29	[35]
miR-501-3p	LIN7A	[36]
miR-874	DOR, EGFR and ERK	[37]
miR-140	MMP9	[38]
Oncogenic miRNAs	Regulated Gene Expressions and/or pathways	
miR-1914	PI3K/Akt	[39]
miR-660-5p	PI3K/Akt	[40]
miR125a-5p	PI3K/Akt	[41]
miR-3131	DTHD1 and XAF1	[42]
miR-122	Induced HCV replication	[43]
miR-17-5p	PTEN, GalNT7, Vimentin	[44]
miR-18a	ESR1	[45]
miR-32	PTEN	[46]
miR-92b	SMAD7	[47]
miR-93	PTEN, CDKN1A, c-Met/p13k/AKT pathway	[48]
miR-101	FOS oncogene	[49]
miR-106b	EMT	[50]
miR-130b	PPAR-γ, TP53INP1	[51]
miR-155	c-Myc, C/EBPβ, APC, β-catenin, Cyclin D1, TP53INP1	[52]
miR-197	Wnt/ $\beta$ -catenin, Axin-2, NKD1 and DKK2	[53]
miR-219-5p	Cadherin 1	[54]
miR-221 and miR-222	PHF2, AKT pathway, PTEN, CDK inhibitor p27 and	[55]
	DDIT4	

Environmental factors, in addition to mutations, have an impact on the mTOR pathway activity. Under hypoxic conditions, HIF-2α was upregulated in steatotic HCC, resulting in activated PI3K and increased levels of phosphorylated AKT, GSK3B and mTOR [26]. In 5–15% of instances, activating mutations of NRF2 (coded by NFE2L2) or inactivating KEAP1 changes the oxidative stress pathway by preventing the proteasome from degrading NRF2, which is physiologically driven by KEAP1/CUL3 complex ubiquitinoylation [56]. Activating the NRF2 pathway has been proven to shield mice from long-term oxidative stress and the development of tumors. NRF2 activation drives tumor growth by recurrent mutations in HCC [57]. *In vitro* studies have shown that activating NRF2 protects tumor cells against ROS exposure, which may otherwise result in death [57].

CNVs and related mutations are prevalent genetic occurrences in HCC. Broad genomic deletions and insertions affecting 1p, 4p-q, 6q, 8p, 13p-q, 16p, 17p, 21p, 22q, and at 1q, 5p, 6p, 8q, 17q, 20q, and Xq have been found [12, 14, 56]. Genes, such as AXIN1, CDKN2A/CDKN2B, CFH, IRF2, MAP2K3, PTEN, PTPN3, RB1, and RPS6KA3 were damaged by recurrent homozygous deletions, whereas high-level focal amplifications affected the loci for VEGFA (1%) and FGF3/4/19/CCND1 (4%) on 6p21 and 11q13, respectively [56, 58]. Fluorescence in situ hybridization was used in further studies to confirm the elevated levels of circulating tumor DNA in these loci [58]. JAK3, MET, and MYC have all been linked to larger DNA increases (3%, 1%, and 1%, respectively) [58]. VEGFA amplifications provide a non-cell autonomous mechanism for sorafenib

sensitiveness [59]. The major signaling pathways in HCC pathophysiology are represented in Figure 1 [60].

#### **Alterations of Metabolic Pathways in HCC**

Metabolic alterations are the common characteristics of cancer cells giving a selective advantage for tumor growth, proliferation, and survival by supplying the critical demands of rapidly growing cancer cells, such as enhanced energy production and accelerated biosynthetic pathways [61]. As the liver is the main site of numerous metabolic events which govern energy metabolism by the regulation of various metabolites including sugars, lipids, and amino acids, it has been indicated that the HCC is accompanied by a highly dysregulated metabolism, compared to normal hepatocytes [62, 63]. Aside from causing metabolic reprogramming, abnormalities in metabolic pathways also remodel the tumor microenvironment through different metabolic signaling pathways, thus synergistically promoting the occurrence and progression of tumors [64]. However, the increased reliance of the tumor on some of these pathways might result in metabolic vulnerabilities in HCC, which can be targeted by inhibitors of these pathways [63]. Therefore, it is crucial to understand the metabolic alterations in HCC cells and the underlying mechanisms behind these alterations on gene and protein levels, as well as their crosstalk with the other signaling pathways, for improving future potential therapeutic interventions.

# Altered Carbohydrate Metabolism in HCC

In physiological conditions, glucose is converted into pyruvate by glycolysis and then completely oxidized in the mitochondria by the tricarboxylic acid (TCA) cycle and the oxidative phosphorylation or can be channeled in the fatty acid synthesis pathway through the *de novo* lipogenesis. However, studies have shown that HCC predominantly displays the Warburg effect with quick energy generation from glycolysis, through the conversion of glucose to lactate, instead of allowing pyruvate to enter the TCA cycle even in the presence of oxygen and fully functioning mitochondria [65]. While glycolysis enzymes are primarily upregulated in HCC, glycogen metabolism enzyme phosphoglucomutase 1 (PGM1) and gluconeogenesis enzymes phosphoenolpyruvate carboxykinases 1 and 2 (PEPCK), and fructose 1,6-bisphosphatase 1 (FBP1) are downregulated, leading glucose to undergo glycolysis instead of being stored as glycogen, and cause loss of the gluconeogenic activity of normal liver cells [63]. Further, HCC tumors enhance glucose uptake by upregulating plasma membrane glucose transporter 1 (GLUT1) and GLUT2 [66, 67]. Accordingly, the TCA cycle is downregulated and due to the increased flux of glycolytic metabolites through the pentose phosphate pathway (PPP), nucleotide metabolism is promoted [68, 69]. The major dysregulated pathways of carbohydrate metabolism and their compounds in HCC are presented in Figure 2 [70]. Overall, this metabolic reprogramming promotes the growth, survival, proliferation, and long-term maintenance of HCC cells, thus, targeting glucose metabolism may serve as a basis for the development of potential drugs in HCC treatment.

# Altered Lipid Metabolism in HCC

The liver is the central organ for fatty acid metabolism, which synthesizes, stores, processes, and breaks down lipids. Therefore, in HCC pathogenesis, several pathways of lipid metabolism are dysregulated, such as fatty acid synthesis,  $\beta$ -oxidation, and cellular lipidic composition, to support tumorigenesis. Studies have shown that the genes involved

in fatty acid and cholesterol biosynthesis are upregulated including mainly acetyl-CoA carboxylase (ACC), fatty-acid synthase (FASN), ATP citrate lyase (ACLY), SCD1 (stearoyl-Coenzyme A desaturase 1), and cholesterol biosynthesis gene 3-hydroxy-3methylalutaryl-CoA reductase (HMGCR), as well as sterol regulatory element-binding protein 1 (SREBP1), an adipogenesis transcriptional regulator [71]. Not only the fatty acid biosynthesis enzymes but also glucose-6-phosphate dehydrogenase (G6PD) and malic enzyme (ME) are upregulated, which produce NADPH supplying the reducing equivalents for fatty acid biosynthesis enzymes [41]. In parallel to the upregulation of fatty acid biosynthesis, the  $\beta$ -oxidation is suppressed by the downregulation of carnitine palmitoyl transferase 2 (CPT2) enzyme expression [41, 72]. Catalyzing the first reaction of ketogenesis, 3-hydroxy-3-methylglutaryl-CoA synthase 2 (HMGCS2) enzyme has also been shown to downregulated in HCC tumor tissues. Lower expression levels of HMGCS2 were correlated with tumor size as well as higher pathological grades, which suggests HMGCS2 may function as a tumor suppressor [62, 73]. However, many metabolites are also associated with various signaling pathways in addition to their roles in energy metabolism. For instance, unsaturated fatty acids have been addressed to downregulate PTEN in hepatocytes through an mTOR/NF-kB-dependent mechanism [74]. Therefore, the crosstalk should be considered in terms of uncovering synergy among pathways and understanding the holistic effects of metabolic components. The major dysregulated pathways of lipid metabolism and their compounds in HCC are presented in Figure 2 [70].

# **Altered Amino Acid Metabolism in HCC**

The amino acid metabolism has been shown to be altered in HCC. Both the amino acids and the related enzymes participate in providing energy and precursors to support tumor initiation and progression. One of the major alterations takes place in aspartate metabolism since the expression levels of asparagine synthetase (ASNS), the enzyme producing asparagine by using aspartate, is decreased in correlation with malignancy of HCC tumors [75]. In HCC, other significant upregulation has been detected in glutamine metabolism with the overexpressed glutamine synthetase (GS) [63]. Despite the increased expression of GS, still, the serum glutamine levels of HCC patients have been found lower than healthy individuals. This outcome is considered the result of increased glutamine uptake by tumors due to upregulated glutamine transporter (ASCT2) [76, 77]. Proline metabolism was also markedly changed in HCC tumor tissue, characterized with accelerated consumption of proline and accumulation of hydroxyproline [78]. However, these amino acids do not only contribute to biosynthesis but also interact with signaling pathways. For instance, glutamine shows its tumor proliferative effects by activating mTORC1 signaling, and accumulated hydroxyproline promotes HCC tumor progression and sorafenib resistance by modulating HIF1 $\alpha$  stability [78]. Additionally, the urea cycle, the process that excretes the byproducts generated by protein degradation, is significantly downregulated in HCC [79]. The major dysregulated steps of urea cycle in HCC are presented in Figure 2 [70]. Overall, these findings imply that targeting the affected metabolic pathways may potentially take an essential part in the HCC therapy. Nevertheless, in HCC pathogenesis, it should be considered that the metabolic pathways crosstalk with each other. Therefore, the synergy of interconnected pathways should be evaluated instead of isolated mechanisms.

# Conclusion

The importance of liver as a vital complex regulatory system for homeostasis is based on a crosstalk between biochemistry and genetics, through the identification of several mutations of genes that impair biochemical pathways and enzyme/protein expressions at different levels with several clinical manifestations. The basic scientific understanding of the biochemical mechanisms those are linked to genetic variants, controlling hepatic tumorigenesis, autoinflammation and autoimmunity has progressed considerably during the past few years, thanks to advances in NGS and GWAS. The challenges for the future will be to understand how the crosstalk between biochemistry and genetics affects the hepatic function and the pathogenesis of liver diseases linked to hepatic tumorigenesis, autoinflammation and autoimmunity. The aim of this review is to update how the crosstalk between genetics and biochemistry affects the pathogenesis of the most common liver diseases. Genetically affected biochemical pathways, both major survival paths of signaling, such as RAF/MEK/ERK, Wnt/B-catenin, PI3K/AKT/mTOR, JAK/STAT, EGFR and MET, and main metabolic paths of biomolecules are evaluated in detail. Some of these important pathways can be novel diagnostic/prognostic biomarkers, and/or novel therapeutic markers for the treatment of these liver diseases. Besides, by unveiling novel genetic variants in hepatic pathogenesis, this review points to the mechanisms that link these genetic variants to impaired biochemical paths, highlighting the importance of several genetic variants and the complexity of biochemical pathways crosstalk in the regulation of hepatocellular homeostasis.

## References

- Llovet, J.M., et al., *Molecular therapies and precision medicine for hepatocellular carcinoma*. Nat Rev Clin Oncol, 2018. 15(10): p. 599-616.
- 2. Gerbes, A., et al., *Gut roundtable meeting paper: selected recent advances in hepatocellular carcinoma*. Gut, 2018. **67**(2): p. 380-388.
- 3. Hoshida, Y., et al., *Molecular classification and novel targets in hepatocellular carcinoma: recent advancements.* Semin Liver Dis, 2010. **30**(1): p. 35-51.
- 4. Villanueva, A., et al., *Combining clinical, pathology, and gene expression data to predict recurrence of hepatocellular carcinoma.* Gastroenterology, 2011. **140**(5): p. 1501-12 e2.
- 5. Boyault, S., et al., *Transcriptome classification of HCC is related to gene alterations and to new therapeutic targets.* Hepatology, 2007. **45**(1): p. 42-52.
- Lachenmayer, A., et al., Wnt-pathway activation in two molecular classes of hepatocellular carcinoma and experimental modulation by sorafenib. Clin Cancer Res, 2012. 18(18): p. 4997-5007.
- Nault, J.C., et al., *The role of telomeres and telomerase in cirrhosis and liver cancer*. Nat Rev Gastroenterol Hepatol, 2019. 16(9): p. 544-558.
- 8. Pezzuto, F., et al., *Frequency and geographic distribution of TERT promoter mutations in primary hepatocellular carcinoma*. Infect Agent Cancer, 2017. **12**: p. 27.
- 9. Buch, S., et al., *Genetic variation in TERT modifies the risk of hepatocellular carcinoma in alcoholrelated cirrhosis: results from a genome-wide case-control study.* Gut, 2023. **72**(2): p. 381-391.
- 10. Wu, R.A., et al., *Telomerase Mechanism of Telomere Synthesis*. Annu Rev Biochem, 2017. 86: p.
  439-460.
- Heidenreich, B. and R. Kumar, *TERT promoter mutations in telomere biology*. Mutat Res Rev Mutat Res, 2017. **771**: p. 15-31.

- 12. Schulze, K., et al., *Exome sequencing of hepatocellular carcinomas identifies new mutational signatures and potential therapeutic targets.* Nat Genet, 2015. **47**(5): p. 505-511.
- Jin, C., et al., Novel copper complex CTB regulates methionine cycle induced TERT
   hypomethylation to promote HCC cells senescence via mitochondrial SLC25A26. Cell Death Dis,
   2020. 11(10): p. 844.
- 14. Totoki, Y., et al., *Trans-ancestry mutational landscape of hepatocellular carcinoma genomes.* Nat Genet, 2014. **46**(12): p. 1267-73.
- 15. Trepo, E., et al., *Common genetic variation in alcohol-related hepatocellular carcinoma: a casecontrol genome-wide association study.* Lancet Oncol, 2022. **23**(1): p. 161-171.
- 16. Calderaro, J., et al., *Molecular and histological correlations in liver cancer*. J Hepatol, 2019. **71**(3):
  p. 616-630.
- Allison, S.J., *Development: beta-catenin network controls nephron patterning*. Nat Rev Nephrol, 2015. 11(4): p. 195.
- Audard, V., et al., Cholestasis is a marker for hepatocellular carcinomas displaying beta-catenin mutations. J Pathol, 2007. 212(3): p. 345-52.
- 19. Bengochea, A., et al., *Common dysregulation of Wnt/Frizzled receptor elements in human hepatocellular carcinoma*. Br J Cancer, 2008. **99**(1): p. 143-50.
- 20. Rana, M.A., et al., *Interplay of Wnt beta-catenin pathway and miRNAs in HBV pathogenesis leading to HCC.* Clin Res Hepatol Gastroenterol, 2019. **43**(4): p. 373-386.
- 21. Klingenberg, M., et al., *Non-coding RNA in hepatocellular carcinoma: Mechanisms, biomarkers and therapeutic targets.* J Hepatol, 2017. **67**(3): p. 603-618.
- 22. Huang, J.L., et al., *Hepatic stellate cells promote the progression of hepatocellular carcinoma through microRNA-1246-RORalpha-Wnt/beta-Catenin axis.* Cancer Lett, 2020. **476**: p. 140-151.

- Xin, R.Q., et al., *MiR-329-3p inhibits hepatocellular carcinoma cell proliferation and migration through USP22-Wnt/beta-Catenin pathway.* Eur Rev Med Pharmacol Sci, 2020. 24(19): p. 9932-9939.
- Cao, X., et al., *microRNA-19a-3p and microRNA-376c-3p Promote Hepatocellular Carcinoma Progression Through SOX6-Mediated Wnt/beta-Catenin Signaling Pathway.* Int J Gen Med, 2021.
   14: p. 89-102.
- 25. Kan, Z., et al., *Whole-genome sequencing identifies recurrent mutations in hepatocellular carcinoma.* Genome Res, 2013. **23**(9): p. 1422-33.
- Villanueva, A., et al., *Pivotal role of mTOR signaling in hepatocellular carcinoma*.
   Gastroenterology, 2008. **135**(6): p. 1972-83, 1983 e1-11.
- Wu, H., et al., *MicroRNA-206 prevents the pathogenesis of hepatocellular carcinoma by modulating expression of met proto-oncogene and cyclin-dependent kinase 6 in mice.* Hepatology, 2017. 66(6): p. 1952-1967.
- 28. Xia, H., L.L. Ooi, and K.M. Hui, *MiR-214 targets beta-catenin pathway to suppress invasion, stemlike traits and recurrence of human hepatocellular carcinoma.* PLoS One, 2012. **7**(9): p. e44206.
- Li, H., H. Wang, and Z. Ren, *MicroRNA-214-5p Inhibits the Invasion and Migration of Hepatocellular Carcinoma Cells by Targeting Wiskott-Aldrich Syndrome Like*. Cell Physiol Biochem, 2018. 46(2): p. 757-764.
- 30. Wong, Q.W., et al., *MicroRNA-223 is commonly repressed in hepatocellular carcinoma and potentiates expression of Stathmin1*. Gastroenterology, 2008. **135**(1): p. 257-69.
- Wang, L., et al., *miRNA-302b suppresses human hepatocellular carcinoma by targeting AKT2*. Mol
   Cancer Res, 2014. 12(2): p. 190-202.
- 32. Sun, J.J., G.Y. Chen, and Z.T. Xie, *MicroRNA-361-5p Inhibits Cancer Cell Growth by Targeting CXCR6 in Hepatocellular Carcinoma.* Cell Physiol Biochem, 2016. **38**(2): p. 777-85.

- 33. Sun, G., et al., *MiR-370 promotes cell death of liver cancer cells by Akt/FoxO3a signalling pathway.* Eur Rev Med Pharmacol Sci, 2016. **20**(10): p. 2011-9.
- 34. He, X.X., et al., *MicroRNA-375 targets AEG-1 in hepatocellular carcinoma and suppresses liver cancer cell growth in vitro and in vivo.* Oncogene, 2012. **31**(28): p. 3357-69.
- 35. Du, H., et al., *MicroRNA-424-5p acts as a potential biomarker and inhibits proliferation and invasion in hepatocellular carcinoma by targeting TRIM29.* Life Sci, 2019. **224**: p. 1-11.
- 36. Luo, C., et al., *microRNA-501-3p suppresses metastasis and progression of hepatocellular carcinoma through targeting LIN7A*. Cell Death Dis, 2018. **9**(5): p. 535.
- 37. Zhang, Y., et al., *microRNA-874 suppresses tumor proliferation and metastasis in hepatocellular carcinoma by targeting the DOR/EGFR/ERK pathway.* Cell Death Dis, 2018. **9**(2): p. 130.
- 38. Lu, Z., et al., *LncRNA FLJ33360 accelerates the metastasis in hepatocellular carcinoma by targeting miRNA-140/MMP9 axis.* Am J Transl Res, 2020. **12**(2): p. 583-591.
- 39. da Silveira, M.B., et al., *The functional activity of the miR-1914-5p in lipid metabolism of the hepatocarcinoma cell line HepG2: a potential molecular tool for controlling hepatic cellular migration.* Mol Biol Rep, 2021. **48**(4): p. 3463-3474.
- 40. Wu, Y., et al., *MiR-660-5p promotes the progression of hepatocellular carcinoma by interaction with YWHAH via PI3K/Akt signaling pathway.* Biochem Biophys Res Commun, 2020. **531**(4): p. 480-489.
- 41. Bjornson, E., et al., *Stratification of Hepatocellular Carcinoma Patients Based on Acetate Utilization.* Cell Rep, 2015. **13**(9): p. 2014-26.
- 42. Wang, C., et al., *An indel polymorphism within pre-miR3131 confers risk for hepatocellular carcinoma.* Carcinogenesis, 2017. **38**(2): p. 168-176.
- 43. Ahsani, Z., et al., WNT1 Gene from WNT Signaling Pathway Is a Direct Target of miR-122 in Hepatocellular Carcinoma. Appl Biochem Biotechnol, 2017. **181**(3): p. 884-897.

- Shan, S.W., et al., Mature miR-17-5p and passenger miR-17-3p induce hepatocellular carcinoma by targeting PTEN, GalNT7 and vimentin in different signal pathways. J Cell Sci, 2013. 126(Pt 6): p. 1517-30.
- 45. Liu, W.H., et al., *MicroRNA-18a prevents estrogen receptor-alpha expression, promoting proliferation of hepatocellular carcinoma cells.* Gastroenterology, 2009. **136**(2): p. 683-93.
- 46. Yan, S.Y., et al., *MiR-32 induces cell proliferation, migration, and invasion in hepatocellular carcinoma by targeting PTEN.* Tumour Biol, 2015. **36**(6): p. 4747-55.
- 47. Zhuang, L.K., et al., *MicroRNA-92b promotes hepatocellular carcinoma progression by targeting Smad7 and is mediated by long non-coding RNA XIST.* Cell Death Dis, 2016. **7**(4): p. e2203.
- 48. Ohta, K., et al., *MicroRNA-93 activates c-Met/PI3K/Akt pathway activity in hepatocellular carcinoma by directly inhibiting PTEN and CDKN1A*. Oncotarget, 2015. **6**(5): p. 3211-24.
- Li, S., et al., *MicroRNA-101 regulates expression of the v-fos FBJ murine osteosarcoma viral oncogene homolog (FOS) oncogene in human hepatocellular carcinoma*. Hepatology, 2009. 49(4):
   p. 1194-202.
- Yau, W.L., et al., Over-expression of miR-106b promotes cell migration and metastasis in hepatocellular carcinoma by activating epithelial-mesenchymal transition process. PLoS One, 2013. 8(3): p. e57882.
- 51. Ma, S., et al., *miR-130b Promotes CD133(+) liver tumor-initiating cell growth and self-renewal via tumor protein 53-induced nuclear protein 1.* Cell Stem Cell, 2010. **7**(6): p. 694-707.
- 52. Higgs, G. and F. Slack, *The multiple roles of microRNA-155 in oncogenesis*. J Clin Bioinforma, 2013. 3(1): p. 17.
- 53. Hu, Z., et al., *MicroRNA-197 Promotes Metastasis of Hepatocellular Carcinoma by Activating Wnt/beta-Catenin Signaling.* Cell Physiol Biochem, 2018. **51**(1): p. 470-486.

- 54. Yang, J., et al., *MicroRNA-219-5p Promotes Tumor Growth and Metastasis of Hepatocellular Carcinoma by Regulating Cadherin 1.* Biomed Res Int, 2018. **2018**: p. 4793971.
- 55. Pineau, P., et al., *miR-221 overexpression contributes to liver tumorigenesis*. Proc Natl Acad Sci U
  S A, 2010. **107**(1): p. 264-9.
- 56. Guichard, C., et al., *Integrated analysis of somatic mutations and focal copy-number changes identifies key genes and pathways in hepatocellular carcinoma.* Nat Genet, 2012. **44**(6): p. 694-8.
- 57. DeNicola, G.M., et al., Oncogene-induced Nrf2 transcription promotes ROS detoxification and tumorigenesis. Nature, 2011. **475**(7354): p. 106-9.
- 58. Sawey, E.T., et al., *Identification of a therapeutic strategy targeting amplified FGF19 in liver cancer by Oncogenomic screening.* Cancer Cell, 2011. **19**(3): p. 347-58.
- 59. Horwitz, E., et al., *Human and mouse VEGFA-amplified hepatocellular carcinomas are highly sensitive to sorafenib treatment.* Cancer Discov, 2014. 4(6): p. 730-43.
- 60. Farzaneh, Z., et al., *Critical signaling pathways governing hepatocellular carcinoma behavior; small molecule-based approaches.* Cancer Cell Int, 2021. **21**(1): p. 208.
- 61. DeBerardinis, R.J., et al., *The biology of cancer: metabolic reprogramming fuels cell growth and proliferation.* Cell Metab, 2008. **7**(1): p. 11-20.
- 62. Huo, J., J. Cai, and L. Wu, *Comprehensive analysis of metabolic pathway activity subtypes derived prognostic signature in hepatocellular carcinoma*. Cancer Med, 2022.
- 63. Tenen, D.G., L. Chai, and J.L. Tan, *Metabolic alterations and vulnerabilities in hepatocellular carcinoma.* Gastroenterol Rep (Oxf), 2021. **9**(1): p. 1-13.
- 64. Lane, A.N., R.M. Higashi, and T.W. Fan, *Metabolic reprogramming in tumors: Contributions of the tumor microenvironment.* Genes Dis, 2020. **7**(2): p. 185-198.
- 65. Chen, H., et al., *Mechanism, Clinical Significance, and Treatment Strategy of Warburg Effect in Hepatocellular Carcinoma.* Journal of Nanomaterials, 2021. **2021**: p. 5164100.

- 66. Kim, Y.H., et al., *SLC2A2 (GLUT2) as a novel prognostic factor for hepatocellular carcinoma.*Oncotarget, 2017. 8(40): p. 68381-68392.
- 67. Amann, T., et al., *GLUT1 expression is increased in hepatocellular carcinoma and promotes tumorigenesis.* Am J Pathol, 2009. **174**(4): p. 1544-52.
- 68. Capuano, F., et al., *Oxidative phosphorylation and F(O)F(1) ATP synthase activity of human hepatocellular carcinoma*. Biochem Mol Biol Int, 1996. **38**(5): p. 1013-22.
- 69. Kowalik, M.A., A. Columbano, and A. Perra, *Emerging Role of the Pentose Phosphate Pathway in Hepatocellular Carcinoma*. Front Oncol, 2017. **7**: p. 87.
- 70. Wang, Q., et al., *Metabolic Reprogramming and Its Relationship to Survival in Hepatocellular Carcinoma*. Cells, 2022. **11**(7).
- 71. Calvisi, D.F., et al., *Increased lipogenesis, induced by AKT-mTORC1-RPS6 signaling, promotes development of human hepatocellular carcinoma*. Gastroenterology, 2011. **140**(3): p. 1071-83.
- 72. Fujiwara, N., et al., *CPT2 downregulation adapts HCC to lipid-rich environment and promotes carcinogenesis via acylcarnitine accumulation in obesity.* Gut, 2018. **67**(8): p. 1493-1504.
- 73. Wang, Y.H., et al., *HMGCS2 Mediates Ketone Production and Regulates the Proliferation and Metastasis of Hepatocellular Carcinoma*. Cancers (Basel), 2019. **11**(12).
- Vinciguerra, M., et al., Unsaturated fatty acids inhibit the expression of tumor suppressor phosphatase and tensin homolog (PTEN) via microRNA-21 up-regulation in hepatocytes.
   Hepatology, 2009. 49(4): p. 1176-84.
- 75. Zhang, B., et al., *Asparagine synthetase is an independent predictor of surgical survival and a potential therapeutic target in hepatocellular carcinoma*. Br J Cancer, 2013. **109**(1): p. 14-23.
- Hirayama, C., et al., *Plasma amino acid patterns in hepatocellular carcinoma*. Biochem Med
   Metab Biol, 1987. **38**(2): p. 127-33.

- 77. Sun, H.W., et al., *GLUT1 and ASCT2 as Predictors for Prognosis of Hepatocellular Carcinoma.* PLoS One, 2016. **11**(12): p. e0168907.
- Tang, L., et al., Global Metabolic Profiling Identifies a Pivotal Role of Proline and Hydroxyproline Metabolism in Supporting Hypoxic Response in Hepatocellular Carcinoma. Clin Cancer Res, 2018.
   24(2): p. 474-485.
- 79. Cao, Y., et al., Significant Down-Regulation of Urea Cycle Generates Clinically Relevant Proteomic Signature in Hepatocellular Carcinoma Patients with Macrovascular Invasion. J Proteome Res, 2019. 18(5): p. 2032-2044.







Fig. 2. The major alterations in metabolic pathways in HCC